Fundamentals of Codon Bias Willow Kion-Crosby¹, Michael Manhart², Unab Javed¹, Manindra Rachakonda¹ and Alex Morozov¹

Introduction

The presence of codon usage bias has been a long-standing mystery [1]. Potential reasons for this bias include,

- Speed of translation.
- Protein synthesis accuracy.



Predictive model

Population genetics model with single-point mutation and absolute fitness. Parameters based on biophysical arguments.

$$|N(t+1)\rangle = (\mathbf{I} + \mathbf{M})\mathbf{W}|N(t)\rangle$$
$$\Rightarrow \langle 1|\mathbf{W}|p_{ss}^c\rangle = (\mathbf{I} + \mathbf{M})\mathbf{W}|p_{ss}^c\rangle.$$

- M, mutational matrix
- W, absolute fitness
- N(t), cell number
- $p^{c}(t)$, codon frequency

After many generations, $t \to \infty$, frequencies reach steady-state p_{ss}^c .



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- [4] F. V. Murphy and V. Ramakrishnan. Structure of a purine-purine wobble base pair in the decoding center of the ribosome. Nat. Struct. Mol. Biol., 11(12):1251–1252, Dec 2004.

¹Department of Physics & Astronomy, Rutgers, The State University of New Jersey ²Department of Chemistry & Chemical Biology, Harvard University

Inference of mutational rates

Obtained through detailed-balance from trimer frequencies.

$$\pi_1 \mu_{21} = \pi_2 \mu_{12} \Rightarrow \begin{cases} \mu_{21} = \beta \kappa \pi_2 \\ \mu_{12} = \beta \kappa \pi_1 \end{cases}$$

Introduces model parameters β , κ_1 and κ_2 corresponding to mutational scale, and transition transversion rate ratios [3].





Protein production rate

Rate of protein production from two mechanisms with effects on fitness captured by a single parameter, T^0 :

- Single mRNA translation rate affects total protein production due to a finite ribosomal reservoir [2].
- tRNA with high cellular concentrations result in faster translation. [1]

Amino acid fidelity & wobble hypothesis

-s penalty for missense mutations and noncognate tRNA binding due to the wobble hypothesis [4].

Wobble rel. rates	
(U at 5' anticodon)	
U/A	$r_{U/A} = 1$
U/G	$r_{U/G} = r_1$
U/U	$r_{U/U} = r_2$
U/C	$r_{U/C} = r_3$



Figure 1: Methionine taking the place of an isoleucine on the protein thrA due to wobble.

cantly

1
(10)
β
κ_1
κ_2
S
T^0
r_1
r_2
r_3

Results

This model has been fit to a 10% training portion of the genome of *Escherichia coli* K-12 MG1655 and tested on the remaining 90%, and the full *Saccharomyces cerevisiae* S288c genome.

Figure 2: Model prediction of codon frequencies in *E. coli*

Observation: frequencies signifiappear depend to mutational "closeness" deleterious codon ON to states (e.g. stop codons.)







Frequencies from S. cerevisiae genome